













Review Article

Redox Mediated Lipid Metabolism in Animals: Mechanisms and Veterinary Relevance in Companion Animals and Wild Birds

Muhammad Hanzalah Yousaf¹ , Umar Aziz¹ , Abdul Rehman² , Muhammad Waqar¹ , Nimra Safdar Ali³ , Nauman Khan¹ , Muhammad Mohsin¹ , Muhammad Yahya Maarij⁴ , Muhammad Mushahid⁵ , and Xiaopeng An^{1*} 

¹ College of Animal Science and Technology, Northwest A&F University, Yangling, China

² Faculty of Animal Production and Technology, Cholistan University of Veterinary & Animal Sciences, Punjab, Pakistan

³ Institute of Animal and Dairy Sciences, Faculty of Animal Husbandry, University of Agriculture Faisalabad, Punjab, Pakistan

⁴ Canterbury Christ Church University, North Holmes Road, Canterbury, England

⁵ University of Agriculture Faisalabad, Constituent College Toba Tek Singh, Toba Tek Singh, Punjab, Pakistan

* **Corresponding author:** Xiaopeng An, College of Animal Science and Technology, Northwest A&F University, Yangling, China. Email: axpdky@nwafu.edu.cn

ARTICLE INFO

Article History:

Received: 12/01/2026

Revised: 24/02/2026

Accepted: 09/03/2026

Published: 31/03/2026



Keywords:

Gut microbiota

Lipid droplet

Lipid metabolism

Metabolic disorder

Oxidative stress

Redox signaling

ABSTRACT

Lipids are essential for animal physiology; however, dysregulated lipid metabolism can induce metabolic stress and impair growth, development, and reproduction. Metabolic homeostasis depends on endocrine-immune system interactions, yet how lipid droplets and organelles, such as the endoplasmic reticulum, mitochondria, lysosomes, and peroxisomes, contribute to stress-induced lipid dysregulation remains unclear. The present study aimed to synthesize current evidence on redox-mediated regulation of lipid metabolism and lipid metabolic disorders in animals, highlighting recent advances, and identify key directions for future studies. The present study summarized evidence on how different dietary lipid classes influence metabolism and animal health, as well as the role of bioactive nutrients in metabolic programming. The current study described the endocrine functions of the liver, gut, and adipose tissue, as well as the stress-related interactions among these organs. The present study indicated how lipid droplets engaged in dynamic organelle interactions during stress progression and evaluated the potential of lipid-focused nutritional interventions as personalized mitigation strategies. In addition, gut microbiota-derived metabolites and related pathways that contribute to redox imbalance, organelle dysfunction, and stress-associated lipid dysregulation were explored. The current study demonstrated that stress-induced disruptions in lipid metabolism involve intricate, multi-organ, and multi-organelle mechanisms driven by redox changes.

1. Introduction

Lipids comprise a diverse class of biomolecules that support animal physiology as major energy substrates, structural components of cellular membranes, and precursors for signaling mediators¹. Key lipid classes include triacylglycerols, cholesterol, phospholipids, and non-esterified fatty acids, each contributing to nutrient absorption, energy balance, hormone synthesis, and cellular communication¹. Disruption of lipid handling can present as dyslipidemia and broader metabolic stress, compromising growth, development, reproductive performance, and immune competence². Metabolic stress related to lipid dysregulation often occurs alongside chronic low-grade

inflammation, indicating coordinated endocrine-immune responses. Hormonal regulators such as insulin, glucagon, leptin, and adiponectin regulate lipid storage, mobilization, and utilization, whereas immune mediators released from metabolic tissues, such as adipose tissue and liver, further influence lipid flux and inflammatory reactions^{3,4}.

Cellular organelles provide the physical and functional platforms that determine lipid fate during health and disease⁵. Lipid droplets store neutral lipids and buffer lipotoxic intermediates by converting excess fatty acids into triacylglycerols and cholesteryl esters⁶. The endoplasmic reticulum (ER) supports lipid synthesis and protein folding,

Cite this paper as: Yousaf MH, Aziz U, Rehman A, Waqar M, Safdar Ali N, Khan N, Mohsin M, Maarij MY, Mushahid M, and An X. Redox Mediated Lipid Metabolism in Animals: Mechanisms and Veterinary Relevance in Companion Animals and Wild Birds. Small Animal Advances. 2026; 5(1): 5-15. DOI: 10.58803/saa.v5i1.47



The Author(s). Published by Rovedar. This is an open-access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

and ER stress can promote triglyceride synthesis⁷, disrupt proteostasis, and intensify inflammatory signaling^{8,9}. Mitochondria generate ATP through fatty-acid β -oxidation and contribute to reactive oxygen species (ROS) production under high substrate flux or respiratory chain strain¹⁰. Peroxisomes participate in very-long-chain fatty-acid oxidation and generate hydrogen peroxide as a by-product, creating a close link between lipid catabolism and redox balance¹¹. Lysosomes play a role in lipid turnover through lipophagy, which is the autophagic process that delivers lipid droplets to lysosomes for breakdown. Impaired lipophagy can lead to longer persistence of lipid droplets and promote ectopic lipid accumulation¹². Disrupted communication among organelles can redirect

fatty acids toward storage rather than oxidation, increase oxidative injury, and accelerate metabolic dysfunction^{13,14}. These alterations emphasized organelle-level mechanisms as essential drivers of stress-linked lipid disorders. **Table 1** provides a summary of organelle-specific contributions to lipid metabolism and redox control, illustrating how impairments in lipid droplets, endoplasmic reticulum, mitochondria, lysosomes, and peroxisomes drive lipotoxicity, inflammation, and metabolic disease progression. The present study aimed to synthesize current knowledge on the role of redox signaling in regulating lipid metabolism within animal disorders and to explore its implications for veterinary health.

Table 1. Key cellular organelles in lipid metabolism and redox regulation in pet animals

Organelle	Primary role in lipid metabolism	Role in redox regulation	Interplay with stress/dysregulation	References
Lipid droplets	Store neutral lipids, including triacylglycerols and cholesteryl esters; buffer lipotoxicity by sequestering excess lipids	Accumulate during stress and help protect cells from lipotoxicity	Under stress conditions, lipid droplets undergo dynamic changes in size and number	(6, 7)
Endoplasmic reticulum	Supports lipid synthesis and protein folding	Contributes to reactive oxygen species production during protein folding	Endoplasmic reticulum stress activates the unfolded protein response and alters lipid synthesis and lipid droplet biogenesis	(8, 12, 13)
Lysosomes	Degrade lipids through lipophagy	Participate in the oxidative degradation of cellular components	Lysosomal dysfunction impairs lipid turnover and promotes lipid accumulation	(2, 4, 6, 15)
Mitochondria	Carry out fatty acid oxidation and ATP production	Major site of reactive oxygen species generation through the electron transport chain	Mitochondrial dysfunction increases reactive oxygen species production and contributes to lipotoxicity and metabolic impairment	(3, 6, 16)
Peroxisomes	Perform beta-oxidation of very long-chain fatty acids and synthesize ether lipids	Generate hydrogen peroxide during fatty acid oxidation and contain antioxidant enzymes	Impaired peroxisomal function reduces fatty acid breakdown and increases oxidative stress	(7, 9, 17)

ATP: Adenosine triphosphate

2. Redox signaling and lipid metabolism

Redox signaling represents a core regulatory system in cells and reflects a dynamic balance between the formation and removal of ROS and reactive nitrogen species (RNS)¹⁸. Although high levels of ROS/RNS can damage biomolecules, controlled ROS/RNS production serves as a signaling mechanism that tunes many cellular functions, including pathways governing lipid metabolism¹⁹. Major ROS include superoxide anion ($O_2^{\bullet-}$), hydrogen peroxide (H_2O_2), and hydroxyl radical ($\bullet OH$), whereas common RNS include nitric oxide ($NO\bullet$) and peroxynitrite ($ONOO^-$)²⁰. Such reactive species are produced during mitochondrial respiration and are generated by enzymes such as nicotinamide adenine dinucleotide phosphate (NADPH) oxidases (NOX), xanthine oxidase, and nitric oxide synthases (NOS). To limit oxidative injury while preserving signaling capacity, cells maintain antioxidant defenses that include enzymatic components (superoxide dismutase, catalase, glutathione peroxidases)²¹ and non-enzymatic buffers (glutathione, thioredoxin, vitamins E and C)²². Redox status, defined by the balance between ROS/RNS production and antioxidant capacity,

shapes the activity of redox-sensitive proteins and signaling pathways that influence lipid handling²³.

Redox regulation affects lipid metabolism at several levels. ROS/RNS can modify the activity of enzymes that control fatty acid synthesis, elongation, desaturation, and oxidation, either directly through oxidative modification or via redox-responsive signaling cascades²⁴. For instance, the cellular redox environment can influence key lipogenic enzymes such as acetyl-CoA carboxylase (ACC) and fatty acid synthase (FAS)²⁵. Fatty-acid degradation pathways, including mitochondrial and peroxisomal β -oxidation, are sensitive to redox conditions because oxidative stress can impair electron transport, limit ATP production, and alter peroxisomal redox outputs. Redox signaling influences lipid metabolism through transcriptional regulation²⁶, including modulation of factors such as peroxisome proliferator-activated receptors (PPARs) and sterol regulatory element-binding proteins (SREBPs), which are sensitive to both nutrient status and cellular redox conditions²⁷. Sustained redox imbalance that results in oxidative stress is closely linked to lipid metabolic disorders in animals, including obesity, hepatic steatosis, and atherosclerotic changes.

Identifying redox-sensitive control points in lipid metabolism may help guide the development of targeted nutritional and therapeutic approaches for the prevention and management of metabolic diseases²⁸.

2.1. Dietary lipids in regulating lipid metabolism and animal health

Dietary lipids supply concentrated energy and provide essential fatty acids and fat-soluble vitamins needed for growth, reproduction, immune competence, and cellular membrane integrity^{29,30}. After intestinal absorption, dietary fatty acids are transported in lipoproteins to tissues for oxidation, storage, or incorporation into structural and signaling lipids³¹. Dietary fatty-acid profiles can influence tissue fatty-acid composition, membrane fluidity, and inflammatory mediator production, thereby shaping metabolic stability and resilience under stress³². Additionally, lipid quality affects redox balance because different fatty acid classes differ in their susceptibility to oxidation and capacity to modulate antioxidant defenses³³.

Fat type plays a major role in oxidative stress and metabolic outcomes³⁴. Diets rich in long-chain omega-3 polyunsaturated fatty acids are commonly linked to lower inflammatory signaling and greater redox balance, partly through enhancement of antioxidant defenses and reduction of excessive pro-inflammatory lipid mediators³⁴⁻³⁶. In small-animal practice, omega-3 supplementation is particularly relevant for inflammatory skin and joint disorders, such as canine atopic dermatitis and feline osteoarthritis, where anti-inflammatory lipid mediators and redox modulation may support clinical improvement³⁷. By contrast, high intake of saturated fats or oxidized lipids can increase lipid peroxidation and disrupt metabolic function, contributing to dyslipidemia, ectopic lipid accumulation, and tissue dysfunction³⁸. A notable clinical example in dogs is the connection between high-fat meals and the risk of pancreatitis, particularly in breeds predisposed to the condition. High-fat diets and hypertriglyceridemia have been linked to pancreatitis in dogs, including Miniature breeds Schnauzers³⁹. These findings emphasized that both the amount and quality of dietary fat can directly influence inflammatory damage and oxidative burden in susceptible animals⁴⁰.

Furthermore, dietary lipids regulate lipid metabolism through nutrient-sensing pathways that adjust lipid uptake, storage, and oxidation in response to substrate availability⁴¹. Omega-3-rich profiles tend to promote fatty acid utilization and anti-inflammatory signaling, whereas excess saturated or oxidized fats tend to lead to lipid accumulation and oxidative stress injury⁴². The balance between omega-6 and omega-3 fatty acids is particularly important because both classes compete for enzymatic pathways that generate lipid mediators with opposing inflammatory effects⁴². Practical diet formulation benefits from careful consideration of the total fat level, the

distribution of fatty acid classes, the management of freshness and oxidation of fats and oils, and alignment with species- and disorder-specific requirements risk. Such precision is essential for minimizing lipid-driven oxidative stress while preserving essential fatty-acid sufficiency and supporting overall health outcomes⁴³.

2.2. Natural products in regulating animal lipid metabolism and disorders

Sources of natural nutrients and bioactive compounds have attracted considerable interest for their possible effects on lipid metabolism regulation and the prevention or alleviation of lipid disorders in animals⁴⁴. The most common modes of action of these compounds include anti-oxidant, anti-inflammatory, and direct metabolic regulating effects⁴⁵. Metabolic programming shows that the benefits of early-life nutrition on metabolic health and susceptibility to affective diseases are long-lasting⁴⁶. Polyphenols present in fruits, vegetables, and available in medicinal plants have a high concentration of potent antioxidant and anti-inflammatory effects⁴⁷. A polyphenol compound found in grapes, resveratrol⁴⁸, was demonstrated to elevate lipid profiles, decrease oxidative stress, and modulate insulin sensitivity in experimental animals with metabolic syndrome⁴⁹. Its action comprises the activation of sirtuin 1 (SIRT1), a protein deacetylase that is significant in controlling metabolism, and the regulation of lipid biosynthetic and catabolic enzymes⁴. Similarly, the active compound in turmeric, curcumin, has demonstrated hypolipidemic properties and antioxidant activity, thereby improving dyslipidaemia and exerting antioxidant effects in cases of induced obesity in animals⁵¹. The actions of curcumin can be attributed to the activation of free radical scavenging, the induction of antioxidant enzymes, and the modulation of signaling pathways via PPARs and nuclear factor kappa B (NF-KB)⁵².

Another group of natural nutrients with a significant impact on lipid metabolism and redox balance is the omega-3 fatty acids, eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA)⁵³. Although these acids are classified as dietary lipids, their potential application in cases of lipid disorders warrants discussion separately⁵⁴. It has been reported that omega-3 fatty acid supplementation can decrease triglyceride and lipoprotein levels, as well as inflammation and oxidative stress, in different animals⁵⁵. These beneficial effects of omega-3 fatty acids are due to their ability to inhibit lipogenesis, promote fatty acid oxidation in the kidneys, and regulate the production of pro-inflammatory agents⁵⁶. Additionally, omega-3 fatty acids may affect gene expression, activating PPAR and thereby positively affecting lipid metabolism and antioxidant defence⁵⁷. Important nutrient classes were discussed in [Table 2](#), highlighting their effects on lipid metabolism, redox control, and key mechanistic pathways.

Table 2. Natural nutrients and their roles in lipid metabolism and redox regulation in small animals and wild birds

Nutrient/product class	Examples	Primary effects on lipid metabolism	Effects on redox regulation	Key mechanisms	References
Polyphenols	Resveratrol, curcumin, quercetin	Improve lipid profiles, reduce lipogenesis, and enhance fatty acid oxidation	Act as antioxidants, scavenge free radicals, and increase antioxidant enzyme activity	SIRT1 activation, NF- κ B modulation, and PPAR-alpha activation	(51, 52)
Omega-3 PUFAs	Eicosapentaenoic acid (EPA), docosahexaenoic acid (DHA)	Lower triglyceride concentrations suppress lipogenesis and promote fatty acid oxidation	Reduce oxidative stress and modulate eicosanoid production	Modulation of inflammatory signaling pathways	(52, 53)
Vitamins	Vitamin E, vitamin C	Support cellular health and indirectly maintain lipid metabolic function	Protect lipids against peroxidation through antioxidant activity	Direct radical scavenging and regeneration of other antioxidants	(54)
Prebiotics	Inulin, fructooligosaccharides	Modulate gut microbiota, increase short-chain fatty acid production, and reduce inflammation	Indirectly improve redox balance through improved gut health	Selective fermentation by beneficial bacteria and production of SCFAs	(55, 56)
Probiotics	<i>Lactobacillus</i> spp., <i>Bifidobacterium</i> spp.	Modulate gut microbiota, improve bile acid metabolism, and reduce inflammation	Indirectly supports redox balance through effects on gut health	Competitive exclusion of pathogens, antimicrobial production, and immune modulation	(56, 57)

NF-KB: Nuclear factor kappa B, PPAR: Peroxisome proliferator-activated receptor, PUFAs: Polyunsaturated fatty acids, SCFAs: Short-chain fatty acids, SIRT1: Sirtuin 1

Other non-pharmaceutical products, including prebiotics and probiotics, indirectly affect lipid metabolism by altering the gut microbiota⁵⁸. It is important to note that prebiotics and probiotics influence natural interventions⁵⁹. Prebiotics and probiotic compounds have the potential to enhance nutrient absorption, modulate bile acid metabolism, and reduce systemic inflammation, thereby improving lipid homeostasis and decreasing oxidative stress. Vitamins, including vitamin E and vitamin C, are also important antioxidants that help protect against severe oxidative damage to lipids and support overall metabolic wellness^{60,61}. The relative shortage of the following vitamins may intensify oxidative stress and contribute to lipid dysregulation⁶². Numerous natural nutrients and products offer opportunities to address animal lipid disorders through metabolic programming. The variety of actions, frequently involving modulation of redox pathways, raises the possibility of nutritional prevention and correction of metabolic diseases in animals⁶³.

3. Endocrine roles of liver, gut, and adipose tissue in stress conditions

The liver, gut, and adipose tissue not only process nutrients but also function as endocrine organs, releasing hormones, cytokines, and metabolites that help regulate overall energy balance and lipid metabolism⁶⁴. Communication among these organs is highly responsive to stress, and stress can disrupt their signaling, increase inflammation, and alter lipid transport and storage⁶⁵. As oxidative burden rises, lipid homeostasis throughout the body becomes increasingly disrupted⁶⁶. The liver plays a central role in controlling lipogenesis, fatty acid oxidation, cholesterol metabolism, and lipoprotein production.

However, during chronic inflammation or prolonged nutrient excess, hepatic lipid metabolism can shift toward triglyceride accumulation and steatosis, rendering the liver more susceptible to oxidative injury and metabolic dysfunction^{67,68}. Moreover, the liver releases endocrine factors, known as hepatokines, such as fibroblast growth factor 21 (FGF21) and fetuin-A, which can influence insulin sensitivity and lipid use in extrahepatic tissues when stress changes their secretion patterns⁶⁹. The gut contributes to endocrine regulation through hormones including glucagon-like peptide-1 (GLP-1), peptide YY (PYY), and cholecystokinin (CCK), which affect appetite, nutrient handling, and insulin secretion⁷⁰. Stress may weaken gut barrier integrity and increase intestinal permeability, allowing microbial products such as lipopolysaccharide and other luminal signals to enter the circulation⁷¹. This can further amplify inflammatory signaling in the liver and adipose tissue, worsen redox imbalance, and promote dyslipidemia and hepatic lipid accumulation⁷². **Figure 1** shows how stress-driven gut dysbiosis and microbial metabolites influence redox signaling, contribute to liver steatosis and adipose inflammation, and may be counteracted by targeted nutritional strategies⁷³. Adipose tissue acts as an active endocrine organ that releases adipokines and inflammatory mediators that influence insulin sensitivity and lipid distribution⁷⁴. Key adipokines include leptin, adiponectin, and resistin, as well as cytokines such as tumor necrosis factor- α (TNF- α) and interleukin-6 (IL-6)⁷⁵. During obesity or chronic inflammatory stress, adipose endocrine output can shift toward a pro-inflammatory profile, contributing to systemic insulin resistance, dyslipidemia, and sustained low-grade inflammation⁷⁶. Communication among the gut, liver, and

adipose tissue occurs through the portal delivery of gut-derived products to the liver and through adipose tissue releasing free fatty acids and adipokines that affect the hepatic metabolism⁶. Stress-activated neuroendocrine signaling, including activation of the hypothalamic-

pituitary-adrenal axis and glucocorticoid elevation, can further promote visceral adiposity, hepatic steatosis, and impaired insulin sensitivity, reinforcing endocrine metabolic dysregulation⁷⁷.

Gut Microbiota → Metabolites → Redox Signaling → Liver/Adipose Lipid Dysregulation (and how nutrition fixes it)

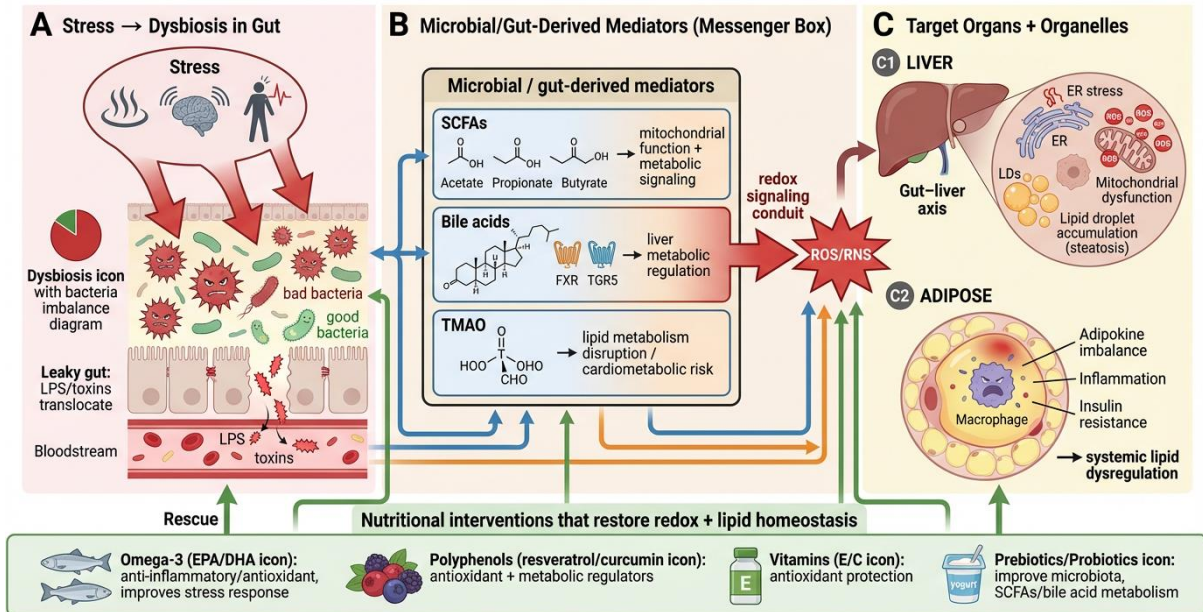


Figure 1. Gut Microbiota, redox, and lipid dysregulation under stress. Under stress, dysbiosis, and increased intestinal permeability allow microbial products and metabolic signals to translocate. These microbial metabolites, including short-chain fatty acids, bile acids, and trimethylamine N-oxide (TMAO), then influence redox signaling and metabolic processes, ultimately affecting hepatic lipid accumulation and adipose inflammation/insulin resistance. Omega-3 fatty acids, polyphenols, vitamins, prebiotics/probiotics can support redox balance and lipid homeostasis. The image was created with PowerPoint and AI (OpenAI, GPT-5.3).

3.1. Response of lipid droplets in stress development

Lipid droplets are dynamic and widespread organelles that function as the main storage sites for neutral lipids, primarily triacylglycerols and cholesteryl esters⁷⁸. Beyond energy storage, lipid droplets contribute to membrane lipid supply, protein sequestration, and signaling. Additionally, lipid droplets provide cytoprotection by buffering excess fatty acids and limiting lipotoxic injury during metabolic stress⁷⁹. Lipid droplet formation is closely associated with the ER. Neutral lipids accumulate between the ER membrane leaflets, then budding occurs to form new lipid droplets in the cytoplasm, which mature by exchanging lipids and proteins with their surrounding organelles⁸⁰.

Lipid droplet abundance and composition change rapidly in response to nutrient limitation, ER stress, and oxidative stress⁸¹. Their formation is often an adaptive mechanism that converts excess free fatty acids into neutral lipids, thereby limiting the buildup of toxic lipid species within the mitochondria and ER. Endoplasmic reticulum stress signaling, including unfolded protein response pathways⁸², can further promote lipid droplet formation by increasing triacylglycerol synthesis and stabilizing ER-lipid droplet contact sites that facilitate lipid transfer transfer⁸³. Interactions between lipid droplets and mitochondria,

lysosomes, and peroxisomes influence cellular stress responses through regulated lipid trafficking. During fasting, lipid droplets can supply fatty acids for β -oxidation, whereas in lipotoxic states, excessive fatty-acid transfer may overwhelm mitochondrial oxidative capacity, increase reactive oxygen species production, and exacerbate cellular injury⁸⁴. Redox state influences lipid droplet dynamics by modulating enzymes involved in triacylglycerol synthesis and lipolysis and by altering organelle contact-site integrity⁸⁵. Oxidative modification of lipid metabolic regulators can alter lipid droplet size and number, while redox-responsive signaling can alter the expression of genes encoding lipid droplet coat proteins and lipid-handling enzymes⁸⁶. Several lipid droplet-associated proteins have mechanistic links between stress signaling and lipid droplet remodeling. These droplets include perilipins (PLIN family), which regulate lipid access at the droplet surface⁸⁷; adipose triglyceride lipase (ATGL/PNPLA2) and hormone-sensitive lipase (HSL/LIPE), which mediate lipolysis, comparative gene identification-58 (CGI-58/ABHD5), serving as an ATGL co-activator, and CIDE family proteins that influence droplet growth and fusion. Under stress conditions, additional functional modules, such as ubiquitin-proteasome components and autophagy-related proteins, are required on the lipid droplet surface. These modules help coordinate

proteostasis and lipid turnover, highlighting a broader role for lipid droplets in cellular adaptation⁸⁸.

Feline hepatic lipidosis is a classic example of how adaptive lipid droplet formation becomes problematic and harmful. Anorexia or extended negative energy balance causes rapid breakdown of adipose tissue and increases the delivery of circulating non-esterified fatty acids to the liver⁸⁹. Hepatocellular esterification converts excess fatty acids into triacylglycerols that accumulate as hepatic lipid droplets. Furthermore, the consistent influx may exceed the liver's capacity for oxidation and export, resulting in an overload of lipid droplets⁵⁹. Lipid droplet expansion is linked to mitochondrial stress, increased reactive oxygen species production, lipid peroxidation, and oxidative damage, further impairing liver function and enhancing appetite suppression. This creates a self-perpetuating cycle of anorexia, lipid mobilization, hepatic lipid droplet buildup, and oxidative stress injury⁹⁰.

Overall, lipid droplet remodeling acts as a coordinated response to nutrient and redox stress, relying on controlled lipid storage⁹¹, mobilization, and inter-organelle communication. Clarifying how lipid droplets interact with organelles and how their associated proteins are regulated under oxidative stress can aid in developing diagnostic markers and nutritional or therapeutic approaches. These strategies aim to reduce lipotoxicity and maintain organelle function in metabolic disorders⁹².

3.2. Nutritional interventions of dietary lipids in response to stress

Dietary lipid planning can lessen stress-associated metabolic dysregulation by reducing lipotoxic exposure, moderating inflammatory signaling⁹², and supporting redox homeostasis. Lipid class (omega-3 versus saturated), lipid oxidation status (fresh versus oxidized fats), and total fat load influence mitochondrial ROS production, lipid peroxidation, and control of lipid synthesis and fatty acid oxidation. Practical applications benefit from choosing specific lipids rather than relying on broad high-fat or low-fat diets, particularly in veterinary contexts where dyslipidemia-related disorders are involved common⁹³.

In small-animal medicine, canine pancreatitis and hypertriglyceridemia-associated pancreatic injury represent clear examples in which dietary lipid control has immediate clinical value⁹⁴. A low-fat, highly digestible diet reduces postprandial triglyceride excursions and limits free fatty acid exposure during lipolysis⁹⁵, thereby lowering inflammatory amplification and oxidative burden. Furthermore, fat quality matters⁹⁶. Omega-3 enrichment (EPA/DHA from fish oil or algal sources) can promote a less pro-inflammatory lipid mediator profile and potentially boost antioxidant defenses through redox-sensitive regulatory pathways, as long as the product quality is maintained and oxidation of supplemental oils is minimized⁹⁷.

In cats, feline hepatic lipidosis underscores the importance of nutritional strategies that restore positive energy balance and limit persistent hepatic lipid accumulation⁹⁸. Core management emphasizes reliable caloric delivery and high-quality protein to reduce peripheral fat mobilization and support hepatic function. Appetite stimulation and assisted feeding plans are often required to achieve consistent intake^{2,99}. Dietary fat should remain moderate rather than extreme, balancing palatability and energy density against the risk of lipotoxicity. Nutrient patterns that promote redox buffering, such as adequate antioxidant micronutrients, can be considered supplementary actions under veterinary supervision, especially when hepatic injury and suspected oxidative stress are present¹⁰. Conjugated linoleic acid can affect body composition and immune signaling, but results vary by isomer and species, limiting its routine clinical use. Medium-chain triglycerides deliver quickly accessible energy and bypass carnitine-dependent mitochondrial transport. These triglycerides may be useful in specific cases, but caution regarding overall fat sensitivity remains important in dogs with pancreatitis³. Overall effectiveness improves when the lipid strategy is combined with complementary measures such as vitamin E and selenium supplementation, microbiome-supportive nutrition (including selected fibers, prebiotics, and probiotics where appropriate), and personalized adjustments based on clinical response and lipid levels⁴.

3.3. Effects of gut microbiota on lipid dysregulation under stress or organelle dysfunction

The gut microbiota, a complex community of microorganisms residing in the gastrointestinal tract, plays a profound role in host metabolism, including lipid homeostasis⁵⁹. This microbiome interacts with the host in multiple ways, such as fermenting food components and generating a diverse array of metabolites that can affect host physiology⁵. Recent findings indicate the influential role of stress in the composition and functions of the gut microbiota, which can subsequently contribute to stress-induced lipid disorders and impairments in lipid-metabolic cell organelles function⁶.

Exposure to stress, whether environmental, psychological, or physiological, can cause dysbiosis, which is a disturbance of the gut microbial balance community⁵⁸. This dysbiosis may result in changes in the metabolic actions of the microbiota, supporting the production of major metabolites, including short-chain fatty acids (SCFAs), bile acids, and other bioactive molecules⁷.

Acetate, propionate, and butyrate are the most common SCFAs produced by dietary fibers and have been demonstrated to play roles in host lipid metabolism as energy providers, in hormone secretion, and in changes in gene expression in metabolic tissues such as the liver and adipose tissue⁸. Stress-related dysbiosis in SCFA production

can mediate the effects of stress-induced dysbiosis on systemic lipid homeostasis.

The bile acids are essential in the digestion and absorption of dietary lipids, which are produced in the liver and metabolized by the gut bacteria⁶⁰. In addition, lipids are signaling molecules that stimulate the nuclear receptors (FXR, farnesoid X) and the G protein-coupled receptor (TGR5) involved in lipid and glucose metabolism, respectively⁹. Changes in gut microbiota due to stress can modulate the deconjugation and transformation of bile acids, and the challenge of changes in the circulating bile acid pool and their resultant effects on host lipid metabolism and intracellular signaling pathways⁷¹. Moreover, intestinal microbiota can produce another substance, trimethylamine n-oxide (TMAO), which has been linked to cardiovascular disease and disrupted lipid metabolism in certain reports¹¹. The gut microbiota or its metabolites may directly or indirectly affect the function of several cell organelles involved in lipid metabolism, such as lipid droplets, ER, mitochondria, lysosomes, and peroxisomes⁶³. For instance, SCFAs have the potential to influence mitochondrial activity and tissue organogenesis¹⁰. Gut microbiota-derived products have the potential to influence ER stress and the unfolded protein response, which are closely linked to lipid metabolism and LD formation¹⁰. This is especially significant to the gut-liver axis, which is a two-way communication between the gut and the liver¹¹. The gut microbiota and the products delivered to the liver through the portal vein directly influence hepatic lipid metabolism and contribute to the development of fatty liver disease stress¹². The redox signal is a vital pathway for communication between the gut microbiota, host tissues, and organelles⁷⁰. Dysbiosis may cause both microbiota and the host immune system in the gut to produce more ROS or RNS, therefore contributing to intestinal oxidative stress. Locally developed oxidative stress can have systemic effects, reflected in the redox status of remote organs involved in lipid metabolism¹³. In addition, gastrointestinal tract microbiota-derived metabolites can directly regulate host antioxidant defense mechanisms and redox-sensitive signaling pathways, and affect lipid homeostasis and organelle activity⁹⁹. Some SCFAs demonstrated antioxidant properties and activated the predicted Nrf2 pathway, a key regulator of antioxidant gene expression¹⁴. The complex interplay between stress and gut microbiota, each with metabolites, redox signaling, and organelle activity, is essential for understanding how stress can be intervened upon in animals to improve lipid dysregulation⁶.

4. Clinical relevance of lipid dysregulation in companion animals

Lifestyle-associated metabolic disease in dogs and cats is increasingly characterized by excess adiposity, dyslipidemia, and chronic low-grade inflammation, with oxidative stress acting as a reinforcing factor for lipid

imbalance⁶⁸. Redox imbalance can increase lipotoxic injury through organelle-level mechanisms, such as mitochondrial ROS production during fatty acid oxidation, endoplasmic reticulum stress responses that promote lipid synthesis and impair protein folding, and maladaptive lipid droplet remodeling that facilitates ectopic lipid accumulation deposition. Common clinical patterns in small-animal practice, including obesity, diabetes phenotypes, hypertriglyceridemia-related pancreatitis risk (especially in dogs), and hepatic lipid disorders (notably feline hepatic lipidosis), all involve mechanisms such as lipid overload, inflammatory responses, and compromised redox balance capacity¹⁶.

Translational management aligns with reducing lipid oversupply and restoring metabolic flexibility². Nutritional strategies include controlled energy intake for weight loss, syndrome-specific macronutrient modifications, and careful management of dietary fat in individuals with hyperlipidemia or pancreatitis dogs⁸⁹. In cats exhibiting insulin-resistance phenotypes, a diet that promotes glycemic stability along with gradual weight loss can help decrease the glucolipotoxic stress on the liver and pancreas. Diet quality is also important, including minimizing oxidized fats and emphasizing lipid sources linked to lower inflammatory signaling, which can support redox balance stability. Adjunctive supplementation offers specific redox support instead of substituting dietary correction. This adjunctive supplementation includes antioxidant nutrients and mitochondrial support, supervised by a veterinarian and tailored to species-specific safety guidelines. Clinical monitoring using body condition scoring, serum triglycerides and cholesterol fractions, and liver-associated enzymes is needed for a greater diet¹⁸.

5. Lipid dysregulation in wild birds

Wild birds in rehabilitation, captivity, or urban areas can develop lipid metabolic issues due to high-energy diets, reduced activity, and chronic stress^{24,63,66}. Obesity, hepatic lipid accumulation, and vascular lipid lesions reported in avian practice are often caused by an imbalance between dietary energy intake and species-appropriate metabolism demand¹⁹. Redox imbalance can contribute to avian lipid pathology by inducing oxidative injury in hepatic tissue, mitochondrial dysfunction during high lipid flux, and inflammatory amplification driven by stress hormones and environmental challenges^{6,20,41}. Such mechanisms align with a redox organelle framework in which sustained oxidative stress disrupts lipid handling and accelerates tissue injury²⁰.

In avian contexts, practical translation mainly focuses on prevention and providing supportive care through proper nutrition husbandry^{27,35,72}. Shifting the diet from energy-dense, seed-heavy patterns to species-appropriate nutrient profiles, combined with foraging enrichment and activity promotion, can reduce lipid overload and improve metabolic health resilience^{24,27,33}. Stress reduction through environmental optimization and minimizing repeated

handling can further limit oxidative burden^{21,42}. In rehabilitation settings, gradual improvement in body condition, careful control of lipid intake during recovery, and regular monitoring of liver function indicators help ensure a safer transition of metabolic processes^{22,48,66}. Nutritional antioxidant support can be considered an adjunct when oxidative injury is suspected, but the focus remains on addressing the primary factors, including diet composition, activity opportunities, and chronic stress exposure, given the strong connection between lifestyle factors and redox-related lipid dysregulation^{22,24}.

6. Conclusion

The present study indicated that redox regulation plays a central role in the pathogenesis of lipid metabolic disorders in animals. The current evidence reflected a complex interplay among cellular pathways, organs, and gut microbiota. These interactions are important to understanding conditions such as obesity, pancreatitis in dogs, fatty liver disease in cats, and similar health issues found in urban wild birds. While the present study supported targeted nutritional and management strategies to mitigate oxidative stress and lipotoxicity, future studies should focus on species-specific biomarkers, controlled dietary intervention trials, and further exploration of organelle interactions and microbiome-derived metabolites to improve diagnosis, treatment, and prevention of diseases, especially in animals.

Declarations

Ethical considerations

Ethical issues, including plagiarism, consent to publish, misconduct, data fabrication and falsification, double publication, submission and redundancy, have been checked by all authors. No AI tool was used to generate scientific content, interpret data, draw conclusions, or create references. The figure was designed by the authors using ChatGPT (OpenAI, GPT-5.3), and the authors take full responsibility for using AI tools for generating the image in the present study.

Funding

No specific funding was received for the present study.

Authors' contributions

Umar Aziz and Muhammad Hanzalah Yousaf conceived and designed the review. Umar Aziz, Abdul Rehman, Muhammad Waqar, and Nimra Safdar Ali performed the literature search and compiled the relevant studies. Umar Aziz drafted the manuscript, and Umar Aziz, Muhammad Waqar, Nauman Khan, and Muhammad Hanzalah Yousaf prepared the figures and visual content. Abdul Rehman, Nimra Safdar Ali, and Muhammad Mushahid critically revised the manuscript for important intellectual content. All authors contributed to writing and editing the review, read and approved the final edition of the manuscript.

Availability of data and materials

No new data were generated in support of the present review. All information is contained within the published literature cited in this article.

Competing interests

The authors declare that they have no competing interests.

References

1. Abou-Rjeileh U, El-Yazbi AF, and El-Sibai M. Redox regulation of lipid mobilization in adipose tissues. *Antioxidants*. 2021; 10(7): 1090. DOI: [10.3390/antiox10071090](https://doi.org/10.3390/antiox10071090)
2. Fransen M, and Lismont C. Redox signaling from and to peroxisomes: Progress, challenges, and prospects. *Antioxid Redox Signal*. 2019; 30(1): 95-112. DOI: [10.1089/ars.2018.7515](https://doi.org/10.1089/ars.2018.7515)
3. Araújo AC, Wheelock CE, and Haeggström JZ. The eicosanoids, redox-regulated lipid mediators in immunometabolic disorders. *Antioxid Redox Signal*. 2018; 29(3): 275-296. DOI: [10.1089/ars.2017.7332](https://doi.org/10.1089/ars.2017.7332)
4. Serrano A, Ribot J, Palou A, Bonet ML. Long-term programming of skeletal muscle and liver lipid and energy metabolism by resveratrol supplementation to suckling mice. *J Nutr Biochem*. 2021; 95: 108770. DOI: [10.1016/j.jnutbio.2021.108770](https://doi.org/10.1016/j.jnutbio.2021.108770)
5. Patias NS, de Queiroz EAlF, Ferrarini SR, Bomfim GF, Aguiar DH, Sinhori AP, et al. Effect of liposomal *Protium heptaphyllum* (Alb.) march extract in the treatment of obesity induced by high-calorie diet. *Biology*. 2024; 13(7): 535. DOI: [10.3390/biology13070535](https://doi.org/10.3390/biology13070535)
6. Chen Z, Tian R, She Z, Cai J, and Li H. Role of oxidative stress in the pathogenesis of nonalcoholic fatty liver disease. *Free Radic Biol Med*. 2020; 152: 116-141. DOI: [10.1016/j.freeradbiomed.2020.02.025](https://doi.org/10.1016/j.freeradbiomed.2020.02.025)
7. Li PL, and Zhang Y. Cross talk between ceramide and redox signaling: Implications for endothelial dysfunction and renal disease. *Handb Exp Pharmacol*. 2013; 216: 171-197. DOI: [10.1007/978-3-7091-1511-4_9](https://doi.org/10.1007/978-3-7091-1511-4_9)
8. Higdon A, Diers AR, Oh JY, Landar A, and Darley-Usmar VM. Cell signalling by reactive lipid species: New concepts and molecular mechanisms. *Biochem J*. 2012; 442(3): 453-464. DOI: [10.1042/BJ20111752](https://doi.org/10.1042/BJ20111752)
9. Niki E. Lipid peroxidation: physiological levels and dual biological effects. *Free Radic Biol Med*. 2009; 47(5): 469-484. DOI: [10.1016/j.freeradbiomed.2009.05.032](https://doi.org/10.1016/j.freeradbiomed.2009.05.032)
10. Forman HJ. Redox signaling: An evolution from free radicals to aging. *Free Radic Biol Med*. 2016; 97: 398-407. DOI: [10.1016/j.freeradbiomed.2016.07.003](https://doi.org/10.1016/j.freeradbiomed.2016.07.003)
11. Yadav UC, and Ramana KV. Regulation of *NF-κB*-induced inflammatory signaling by lipid peroxidation-derived aldehydes. *Oxid Med Cell Longev*. 2013; 2013: 690545. DOI: [10.1155/2013/690545](https://doi.org/10.1155/2013/690545)
12. Distéfano AM, López GA, Bauer V, Zabaleta E, and Pagnussat GC. Ferroptosis in plants: Regulation of lipid peroxidation and redox status. *Biochem J*. 2022; 479(7): 857-866. DOI: [10.1042/BCJ20210682](https://doi.org/10.1042/BCJ20210682)
13. Clérin E, Ait-Ali N, Sahel JA, and Léveillard T. Restoration of rod-derived metabolic and redox signaling to prevent blindness. *Cold Spring Harb Perspect Med*. 2024; 14(11): a041284. DOI: [10.1101/cshperspect.a041284](https://doi.org/10.1101/cshperspect.a041284)
14. Ushio-Fukai M. Compartmentalization of redox signaling through NADPH oxidase-derived ROS. *Antioxid Redox Signal*. 2009; 11(6): 1289-1299. DOI: [10.1089/ars.2008.2333](https://doi.org/10.1089/ars.2008.2333)
15. Circu ML, and Aw TY. Reactive oxygen species, cellular redox systems, and apoptosis. *Free Radic Biol Med*. 2010; 48(6): 749-762. DOI: [10.1016/j.freeradbiomed.2009.12.022](https://doi.org/10.1016/j.freeradbiomed.2009.12.022)
16. Touyz RM. Reactive oxygen species, vascular oxidative stress, and redox signaling in hypertension: What is the clinical significance? *Hypertension*. 2004; 44(3): 248-252. DOI: [10.1161/01.HYP.0000138070.47616.9d](https://doi.org/10.1161/01.HYP.0000138070.47616.9d)
17. Brigelius-Flohé R, and Flohé LI. Basic principles and emerging concepts in the redox control of transcription factors. *Antioxid Redox Signal*. 2011; 15(8): 2335-2381. DOI: [10.1089/ars.2010.3534](https://doi.org/10.1089/ars.2010.3534)
18. Morgan MJ, and Liu ZG. Lipid rafts and oxidative stress-induced cell death. *Antioxid Redox Signal*. 2007; 9(9): 1471-1483. DOI: [10.1089/ars.2007.1658](https://doi.org/10.1089/ars.2007.1658)

19. Ježek P. Physiological fatty acid-stimulated insulin secretion and redox signaling versus lipotoxicity. *Antioxid Redox Signal.* 2025; 42(10-12): 566-622. DOI: [10.1089/ars.2024.0799](https://doi.org/10.1089/ars.2024.0799)
20. Thomas SR, Maiocchi S, Taylor WR, Cai H, and Thomas DD. Redox control of endothelial function and dysfunction: Molecular mechanisms and therapeutic opportunities. *Antioxid Redox Signal.* 2008; 10(10): 1713-1765. DOI: [10.1089/ars.2008.2027](https://doi.org/10.1089/ars.2008.2027)
21. Tai CC, and Ding ST. N-3 polyunsaturated fatty acids regulate lipid metabolism through several inflammation mediators: Mechanisms and implications for obesity prevention. *J Nutr Biochem.* 2010; 21(5): 357-363. DOI: [10.1016/j.jnutbio.2009.09.010](https://doi.org/10.1016/j.jnutbio.2009.09.010)
22. Bergen WG, and Mersmann HJ. Comparative aspects of lipid metabolism: Impact on contemporary research and use of animal models. *J Nutr.* 2005; 135(11): 2499-2502. DOI: [10.1093/jn/135.11.2499](https://doi.org/10.1093/jn/135.11.2499)
23. Ringseis R, and Eder K. Regulation of genes involved in lipid metabolism by dietary oxidized fat. *Mol Nutr Food Res.* 2011; 55(1): 109-121. DOI: [10.1002/mnfr.201000424](https://doi.org/10.1002/mnfr.201000424)
24. Bergen WG, and Brandebourg TD. Regulation of lipid deposition in farm animals: Parallels between agriculture and human physiology. *Exp Biol Med (Maywood).* 2016; 241(12): 1272-1280. DOI: [10.1177/1535370216654996](https://doi.org/10.1177/1535370216654996)
25. Wang YM, Zhang B, Xue Y, Li ZJ, Wang JF, Xue CH, et al. The mechanism of dietary cholesterol effects on lipids metabolism in rats. *Lipids Health Dis.* 2010; 9: 4. DOI: [10.1186/1476-511X-9-4](https://doi.org/10.1186/1476-511X-9-4)
26. Hulbert AJ, Turner N, Storlien LH, and Else PL. Dietary fats and membrane function: Implications for metabolism and disease. *Biol Rev Camb Philos Soc.* 2005; 80(1): 155-169. DOI: [10.1017/s1464793104006578](https://doi.org/10.1017/s1464793104006578)
27. Ding X, Giannenas I, Skoufos I, Wang J, and Zhu W. The effects of plant extracts on lipid metabolism of chickens – A review. *Anim Biosci.* 2023; 36(5): 679-691. DOI: [10.5713/ab.22.0272](https://doi.org/10.5713/ab.22.0272)
28. Ko CW, Qu J, Black DD, and Tso P. Regulation of intestinal lipid metabolism: Current concepts and relevance to disease. *Nat Rev Gastroenterol Hepatol.* 2020; 17(3): 169-183. DOI: [10.1038/s41575-019-0250-7](https://doi.org/10.1038/s41575-019-0250-7)
29. Zhang W, Dan Z, Zheng J, Du J, Liu Y, Zhao Z, et al. Optimal dietary lipid levels alleviated adverse effects of high temperature on growth, lipid metabolism, antioxidant and immune responses in juvenile turbot (*Scophthalmus maximus* L.). *Comp Biochem Physiol B Biochem Mol Biol.* 2024; 272: 110962. DOI: [10.1016/j.cbpb.2024.110962](https://doi.org/10.1016/j.cbpb.2024.110962)
30. Scollan ND, Dannenberger D, Nuernberg K, Richardson I, MacKintosh S, Hocquette JF, et al. Enhancing the nutritional and health value of beef lipids and their relationship with meat quality. *Meat Sci.* 2014; 97(3): 384-394. DOI: [10.1016/j.meatsci.2014.02.015](https://doi.org/10.1016/j.meatsci.2014.02.015)
31. Jin A, Kan Z, Tan Q, Shao J, Han Q, Chang Y, et al. Supplementation with food-derived oligopeptides promotes lipid metabolism in young male cyclists: A randomized controlled crossover trial. *J Int Soc Sports Nutr.* 2023; 20(1): 2254741. DOI: [10.1080/15502783.2023.2254741](https://doi.org/10.1080/15502783.2023.2254741)
32. Chilliard Y, and Ferlay A. Dietary lipids and forages interactions on cow and goat milk fatty acid composition and sensory properties. *Reprod Nutr Dev.* 2004; 44(5): 467-492. DOI: [10.1051/rnd:2004052](https://doi.org/10.1051/rnd:2004052)
33. Navidshad B, and Royan M. Effect of dietary fat on gene expression in poultry, a review. *Crit Rev Eukaryot Gene Expr.* 2016; 26(4): 333-341. DOI: [10.1615/CritRevEukaryotGeneExpr.2016016859](https://doi.org/10.1615/CritRevEukaryotGeneExpr.2016016859)
34. Zhang L, Li Y, Guo Q, Duan Y, Wang W, Zhong Y, et al. Leucine supplementation: A novel strategy for modulating lipid metabolism and energy homeostasis. *Nutrients.* 2020; 12(5): 1299. DOI: [10.3390/nu12051299](https://doi.org/10.3390/nu12051299)
35. Kouba M, and Mourot J. A review of nutritional effects on fat composition of animal products with special emphasis on n-3 polyunsaturated fatty acids. *Biochimie.* 2011; 93(1): 13-17. DOI: [10.1016/j.biochi.2010.02.027](https://doi.org/10.1016/j.biochi.2010.02.027)
36. Kuhla B, Metges CC, and Hammon HM. Endogenous and dietary lipids influencing feed intake and energy metabolism of periparturient dairy cows. *Domest Anim Endocrinol.* 2016; 56: S2-S10. DOI: [10.1016/j.domaniend.2015.12.002](https://doi.org/10.1016/j.domaniend.2015.12.002)
37. Liu Y, Zhang H, Yang X, and Xie C. Dietary uridine improves lipid homeostasis in high-fat diet-induced obese mice by regulating liver gene expression and metabolic profiles. *Front Nutr.* 2025; 12: 1651993. DOI: [10.3389/fnut.2025.1651993](https://doi.org/10.3389/fnut.2025.1651993)
38. Vasantha Rupasinghe HP, Sekhon-Loodu S, Mantso T, and Panayiotidis MI. Phytochemicals in regulating fatty acid β -oxidation: Potential underlying mechanisms and their involvement in obesity and weight loss. *Pharmacol Ther.* 2016; 165: 153-163. DOI: [10.1016/j.pharmthera.2016.06.005](https://doi.org/10.1016/j.pharmthera.2016.06.005)
39. Luo Y, Cheng R, Liang H, Miao Z, Wang J, Zhou Q, et al. Influence of high-fat diet on host animal health via bile acid metabolism and benefits of oral-fed *Streptococcus thermophilus* MN-ZLW-002. *Exp Anim.* 2022; 71(4): 468-480. DOI: [10.1538/expanim.21-0182](https://doi.org/10.1538/expanim.21-0182)
40. Jump DB, and Clarke SD. Regulation of gene expression by dietary fat. *Annu Rev Nutr.* 1999; 19: 63-90. DOI: [10.1146/annurev.nutr.19.1.63](https://doi.org/10.1146/annurev.nutr.19.1.63)
41. Yan K. Recent advances in the effect of adipose tissue inflammation on insulin resistance. *Cell Signal.* 2024; 120: 111229. DOI: [10.1016/j.cellsig.2024.111229](https://doi.org/10.1016/j.cellsig.2024.111229)
42. Beaupere C, Liboz A, Fève B, Blondeau B, and Guillemain G. Molecular mechanisms of glucocorticoid-induced insulin resistance. *Int J Mol Sci.* 2021; 22(2): 623. DOI: [10.3390/ijms22020623](https://doi.org/10.3390/ijms22020623)
43. Singh V, Mahra K, Jung D, and Shin JH. Gut microbes in polycystic ovary syndrome and associated comorbidities; type 2 diabetes, non-alcoholic fatty liver disease (NAFLD), cardiovascular disease (CVD), and the potential of microbial therapeutics. *Probiotics Antimicrob Proteins.* 2024; 16(5): 1744-1761. DOI: [10.1007/s12602-024-10262-y](https://doi.org/10.1007/s12602-024-10262-y)
44. Sarwar H, Rafiqi SI, Ahmad S, Jinna S, Khan SA, Karim T, et al. Hyperinsulinemia associated depression. *Clin Med Insights Endocrinol Diabetes.* 2022; 15: 11795514221090244. DOI: [10.1177/11795514221090244](https://doi.org/10.1177/11795514221090244)
45. Zhang YJ, Gan RY, Li S, Zhou Y, Li AN, Xu DP, et al. Antioxidant Phytochemicals for the prevention and treatment of chronic diseases. *molecules.* 2015; 20(12): 21138-21156. DOI: [10.3390/molecules201219753](https://doi.org/10.3390/molecules201219753)
46. Kilwein MD, Dao TK, and Welte MA. *Drosophila* embryos allocate lipid droplets to specific lineages to ensure punctual development and redox homeostasis. *PLoS Genet.* 2023; 19(8): e1010875. DOI: [10.1371/journal.pgen.1010875](https://doi.org/10.1371/journal.pgen.1010875)
47. Jarc E, and Petan T. A twist of FATE: Lipid droplets and inflammatory lipid mediators. *Biochimie.* 2020; 169: 69-87. DOI: [10.1016/j.biochi.2019.11.016](https://doi.org/10.1016/j.biochi.2019.11.016)
48. de Andrade Melo-Sterza F, and Poehland R. Lipid metabolism in bovine oocytes and early embryos under *in vivo*, *in vitro*, and stress conditions. *Int J Mol Sci.* 2021; 22(7): 3421. DOI: [10.3390/ijms22073421](https://doi.org/10.3390/ijms22073421)
49. Tan Y, Jin Y, Wang Q, Huang J, Wu X, and Ren Z. Perilipin 5 protects against cellular oxidative stress by enhancing mitochondrial function in HepG2 cells. *Cells.* 2019; 8(10): 1241. DOI: [10.3390/cells8101241](https://doi.org/10.3390/cells8101241)
50. Hussain SS, Tran TM, Ware TB, Luse MA, Prevost CT, Ferguson AN, et al. RALA and PLD1 promote lipid droplet growth in response to nutrient withdrawal. *Cell Rep.* 2021; 36(4): 109451. DOI: [10.1016/j.celrep.2021.109451](https://doi.org/10.1016/j.celrep.2021.109451)
51. Nerstedt A, Kurhe Y, Cansby E, Caputo M, Gao L, Vorontsov E, et al. Lipid droplet-associated kinase STK25 regulates peroxisomal activity and metabolic stress response in steatotic liver. *J Lipid Res.* 2020; 61(2): 178-191. DOI: [10.1194/jlr.RA119000316](https://doi.org/10.1194/jlr.RA119000316)
52. Ralhan I, Chang CL, Lippincott-Schwartz J, and Ioannou MS. Lipid droplets in the nervous system. *Lipid droplets in the nervous system.* *J Cell Biol.* 2021; 220(7): e202102136. DOI: [10.1083/jcb.202102136](https://doi.org/10.1083/jcb.202102136)
53. VandeKopple MJ, Wu J, Auer EN, Giaccia AJ, Denko NC, and Papandreou I. HILPDA regulates lipid metabolism, lipid droplet abundance, and response to microenvironmental stress in solid tumors. *Mol Cancer Res.* 2019; 17(10): 2089-2101. DOI: [10.1158/1541-7786.MCR-18-1343](https://doi.org/10.1158/1541-7786.MCR-18-1343)
54. Teixeira PG, David F, Siewers V, and Nielsen J. Engineering lipid droplet assembly mechanisms for improved triacylglycerol accumulation in *Saccharomyces cerevisiae*. *FEMS Yeast Res.* 2018; 18(6): foy060. DOI: [10.1093/femsyr/foy060](https://doi.org/10.1093/femsyr/foy060)
55. Wei C, Yan Y, Miao X, and Jiao R. Dissection and lipid droplet staining of oenocytes in *Drosophila* larvae. *J Vis Exp.* 2019; (154): e60606. DOI: [10.3791/60606](https://doi.org/10.3791/60606)
56. Rimkus SA, Ganetzky B, and Wassarman DA. Traumatic brain injury reprograms lipid droplet metabolism shaped by aging and diet in *Drosophila* brain. *PLoS One.* 2025; 20(9): e0332333. DOI: [10.1371/journal.pone.0332333](https://doi.org/10.1371/journal.pone.0332333)
57. Bradley J, and Swann K. Mitochondria and lipid metabolism in mammalian oocytes and early embryos. *Int J Dev Biol.* 2019; 63(3-4-5): 93-103. DOI: [10.1387/ijdb.180355ks](https://doi.org/10.1387/ijdb.180355ks)
58. Asimakopoulou A, Borkham-Kamphorst E, Henning M, Yagmur E, Gassler N, Liedtke C, et al. Lipocalin-2 (LCN2) regulates PLIN5 expression and intracellular lipid droplet formation in the liver. *Biochim Biophys Acta.* 2014; 1842(10): 1513-1524. DOI: [10.1016/j.bba.2014.08.011](https://doi.org/10.1016/j.bba.2014.08.011)

- 10.1016/j.bbali.2014.07.017
59. Li M, Zhao B, Wang J, Zhang H, Yang Y, Song S, et al. Caveolin 1 in bovine liver is associated with fatty acid-induced lipid accumulation and the endoplasmic reticulum unfolded protein response: Role in fatty liver development. *J Dairy Sci.* 2025; 108(1): 1007-1021. DOI: [10.3168/jds.2024-25349](https://doi.org/10.3168/jds.2024-25349)
 60. Ackerman D, Tumanov S, Qiu B, Michalopoulou E, Spata M, Azzam A, et al. Triglycerides promote lipid homeostasis during hypoxic stress by balancing fatty acid saturation. *Cell Rep.* 2018; 24(10): 2596-2605. DOI: [10.1016/j.celrep.2018.08.015](https://doi.org/10.1016/j.celrep.2018.08.015)
 61. Li Y, De J, Jiang Q, Yang Y, Xu W, Du X, et al. Comparison of lipid metabolism between broodstock and hybrid offspring in the hepatopancreas of juvenile shrimp (*Macrobrachium nipponense*): Response to chronic ammonia stress. *Anim Genet.* 2022; 53(3): 393-404. DOI: [10.1111/age.13194](https://doi.org/10.1111/age.13194)
 62. Fueser H, Majidi N, Haegerbaeumer A, Pilger C, Hachmeister H, Greife P, et al. Analyzing life-history traits and lipid storage using CARS microscopy for assessing effects of copper on the fitness of *Caenorhabditis elegans*. *Ecotoxicol Environ Saf.* 2018; 156: 255-262. DOI: [10.1016/j.ecoenv.2018.03.037](https://doi.org/10.1016/j.ecoenv.2018.03.037)
 63. Abbink MR, Schipper L, Naninck EFG, de Vos CMH, Meier R, van Der Beek EM, et al. The effects of early life stress, postnatal diet modulation, and long-term Western-style diet on later-life metabolic and cognitive outcomes. *Nutrients.* 2020; 12(2): 570. DOI: [10.3390/nu12020570](https://doi.org/10.3390/nu12020570)
 64. Bisogno S, Depciuch J, Gulzar H, Heber MF, Kobińska M, Gąsior Ł, et al. Female-age-dependent changes in the lipid fingerprint of the mammalian oocytes. *Hum Reprod.* 2024; 39(12): 2754-2767. DOI: [10.1093/humrep/deae225](https://doi.org/10.1093/humrep/deae225)
 65. Wang L, Lin J, Yu J, Yang K, Sun L, Tang H, et al. Downregulation of Perilipin1 by the Immune deficiency pathway leads to lipid droplet reconfiguration and adaptation to bacterial infection in *Drosophila*. *J Immunol.* 2021; 207(9): 2347-2358. DOI: [10.4049/jimmunol.2100343](https://doi.org/10.4049/jimmunol.2100343)
 66. Min L, Li D, Tong X, Nan X, Ding D, Xu B, et al. Nutritional strategies for alleviating the detrimental effects of heat stress in dairy cows: A review. *Int J Biometeorol.* 2019; 63(9): 1283-1302. DOI: [10.1007/s00484-019-01744-8](https://doi.org/10.1007/s00484-019-01744-8)
 67. Sordillo LM, and Raphael W. Significance of metabolic stress, lipid mobilization, and inflammation on transition cow disorders. *Vet Clin North Am Food Anim Pract.* 2013; 29(2): 267-278. DOI: [10.1016/j.cvfa.2013.03.002](https://doi.org/10.1016/j.cvfa.2013.03.002)
 68. Mohammed BM, Mohamed Ahmed IA, Alshammari GM, Osman MA, and Yahya MA. Fermented and germinated Samh seeds reduces hyperlipidemia, oxidative stress, and inflammation in rats fed a high-fat diet. *Mol Nutr Food Res.* 2025; 69(21): e70204. DOI: [10.1002/mnfr.70204](https://doi.org/10.1002/mnfr.70204)
 69. Sanchez-Roman I, and Barja G. Regulation of longevity and oxidative stress by nutritional interventions: Role of methionine restriction. *Exp Gerontol.* 2013; 48(10): 1030-1042. DOI: [10.1016/j.exger.2013.02.021](https://doi.org/10.1016/j.exger.2013.02.021)
 70. Popović T, Borozan S, Arsić A, Martačić JD, Vučić V, Trbović A, et al. Fish oil supplementation improved liver phospholipids fatty acid composition and parameters of oxidative stress in male Wistar rats. *J Anim Physiol Anim Nutr (Berl).* 2012; 96(6): 1020-1029. DOI: [10.1111/j.1439-0396.2011.01216.x](https://doi.org/10.1111/j.1439-0396.2011.01216.x)
 71. Yu BP, LIM BO, and SUGANO M. Dietary restriction downregulates free radical and lipid peroxide production: Plausible mechanism for elongation of life span. *J Nutr Sci Vitaminol (Tokyo).* 2002; 48(4): 257-264. DOI: [10.3177/jnsv.48.257](https://doi.org/10.3177/jnsv.48.257)
 72. Ponnampalam EN, Kiani A, Santhiravel S, Holman BWB, Lauridsen C, and Dunshea FR. The importance of dietary antioxidants on oxidative stress, meat and milk production, and their preservative aspects in farm animals: Antioxidant action, animal health, and product quality – Invited Review. *Animals.* 2022; 12(23): 3279. DOI: [10.3390/ani12233279](https://doi.org/10.3390/ani12233279)
 73. Gao J, Ren H, Wu X, Zou C, He B, and Ma W. Dietary glycerol monolaurate mitigates heat stress-induced disruption of intestinal homeostasis and hepatic lipid metabolism in laying hens. *Stress Biol.* 2025; 5(1): 49. DOI: [10.1007/s44154-025-00243-8](https://doi.org/10.1007/s44154-025-00243-8)
 74. Qi M, Wang J, Tan B, Li J, Liao S, Liu Y, and Yin Y. Dietary glutamine, glutamate, and aspartate supplementation improves hepatic lipid metabolism in post-weaning piglets. *Anim Nutr.* 2020; 6(2): 124-129. DOI: [10.1016/j.aninu.2019.12.002](https://doi.org/10.1016/j.aninu.2019.12.002)
 75. Dasilva G, Pazos M, Garcia-Egido E, Pérez-Jiménez J, Torres JL, Giralt MJ, et al. Lipidomics to analyze the influence of diets with different EPA:DHA ratios in the progression of metabolic syndrome using SHROB rats as a model. *Food Chem.* 2016; 205: 196-203. DOI: [10.1016/j.foodchem.2016.03.020](https://doi.org/10.1016/j.foodchem.2016.03.020)
 76. Hunsche C, de Toda IM, Hernandez O, Jiménez B, Díaz LE, Marcos A, et al. The supplementations with 2-hydroxyoleic acid and n-3 polyunsaturated fatty acids revert oxidative stress in various organs of diet-induced obese mice. *Free Radic Res.* 2020; 54(6): 455-466. DOI: [10.1080/10715762.2020.1800004](https://doi.org/10.1080/10715762.2020.1800004)
 77. Lee J, Lee JK, Lee JJ, Park S, Jung S, Lee HJ, et al. Partial replacement of high-fat diet with beef tallow attenuates dyslipidemia and endoplasmic reticulum stress in *db/db* mice. *J Med Food.* 2022; 25(6): 660-674. DOI: [10.1089/jmf.2022.K.0019](https://doi.org/10.1089/jmf.2022.K.0019)
 78. Rincón-Cervera MA, Valenzuela R, Hernández-Rodas MC, Marambio M, Espinosa A, Mayer S, et al. Supplementation with antioxidant-rich extra virgin olive oil prevents hepatic oxidative stress and reduction of desaturation capacity in mice fed a high-fat diet: Effects on fatty acid composition in liver and extrahepatic tissues. *Nutrition.* 2016; 32(11-12): 1254-1267. DOI: [10.1016/j.nut.2016.04.006](https://doi.org/10.1016/j.nut.2016.04.006)
 79. Liu H, Nie C, Hu X, and Li J. Highland barley β -glucan supplementation attenuated hepatic lipid accumulation in Western diet-induced non-alcoholic fatty liver disease mice by modulating gut microbiota. *Food Funct.* 2024; 15(3): 1250-1264. DOI: [10.1039/d3fo03386d](https://doi.org/10.1039/d3fo03386d)
 80. Dong Y, Li L, Xia T, Wang L, Xiao L, Ding N, et al. Oxidative stress can be attenuated by 4-PBA caused by high-fat or ammonia nitrogen in cultured spotted seabass: The mechanism is related to endoplasmic reticulum stress. *Antioxidants.* 2022; 11(7): 1276. DOI: [10.3390/antiox11071276](https://doi.org/10.3390/antiox11071276)
 81. Yanguas-Casás N, Torres-Fuentes C, Crespo-Castrillo A, Diaz-Pacheco S, Healy K, Stanton CS, et al. High-fat diet alters stress behavior, inflammatory parameters and gut microbiota in Tg APP mice in a sex-specific manner. *Neurobiol Dis.* 2021; 159: 105495. DOI: [10.1016/j.nbd.2021.105495](https://doi.org/10.1016/j.nbd.2021.105495)
 82. Chu MM, Luyer MDP, Wheelhouse NM, Bellamy CO, Greve JWM, Buurman WA, et al. Effect of high-fat enteral nutrition on hepatocyte injury in response to hemorrhagic shock in the rat. *World J Surg.* 2007; 31(8): 1693-1701. DOI: [10.1007/s00268-007-9107-2](https://doi.org/10.1007/s00268-007-9107-2)
 83. Khan MZ, Huang B, Kou X, Chen Y, Liang H, Ullah Q, et al. Enhancing bovine immune, antioxidant and anti-inflammatory responses with vitamins, rumen-protected amino acids, and trace minerals to prevent periparturient mastitis. *Front Immunol.* 2024; 14: 1290044. DOI: [10.3389/fimmu.2023.1290044](https://doi.org/10.3389/fimmu.2023.1290044)
 84. Santamarina AB, Sertorio MN, Mennitti LV, de Souza EA, de Souza DV, Ribeiro DA, et al. Hepatic effects of low-carbohydrate diet associated with different lipid sources: Insights into oxidative stress, cytotoxicity, and epigenetic markers in a mouse model of obesity. *J Nutr.* 2024; 154(5): 1517-1531. DOI: [10.1016/j.tjn.2024.04.007](https://doi.org/10.1016/j.tjn.2024.04.007)
 85. Most MS, and Yates DT. Inflammatory mediation of heat stress-induced growth deficits in livestock and its potential role as a target for nutritional interventions: A review. *Animals.* 2021; 11(12): 3539. DOI: [10.3390/ani11123539](https://doi.org/10.3390/ani11123539)
 86. Wang L, Yang T, Pan Y, Shi L, Jin Y, and Huang X. The metabolism of reactive oxygen species and their effects on lipid biosynthesis of microalgae. *Int J Mol Sci.* 2023; 24(13): 11041. DOI: [10.3390/ijms241311041](https://doi.org/10.3390/ijms241311041)
 87. Liu JL, and Hekimi S. The impact of mitochondrial oxidative stress on bile acid-like molecules in *C. elegans* provides a new perspective on human metabolic diseases. *Worm.* 2013; 2(1): e21457. DOI: [10.4161/worm.21457](https://doi.org/10.4161/worm.21457)
 88. Wang W, Gao X, Liu L, Guo S, Duan J, and Xiao P. Zebrafish as a vertebrate model for high-throughput drug toxicity screening: Mechanisms, novel techniques, and future perspectives. *J Pharm Anal.* 2025; 15(9): 101195. DOI: [10.1016/j.jpha.2025.101195](https://doi.org/10.1016/j.jpha.2025.101195)
 89. Min L, Zhao S, Tian H, Zhou X, Zhang Y, Li S, et al. Metabolic responses and “omics” technologies for elucidating the effects of heat stress in dairy cows. *Int J Biometeorol.* 2017; 61(6): 1149-1158. DOI: [10.1007/s00484-016-1283-z](https://doi.org/10.1007/s00484-016-1283-z)
 90. Rehman ZU, Meng C, Yingjie S, Safdar A, Pasha RH, Munir M, et al. Oxidative stress in poultry: Lessons from the viral infections. *Oxid Med Cell Longev.* 2018; 2018(1): 5123147. DOI: [10.1155/2018/5123147](https://doi.org/10.1155/2018/5123147)
 91. Karnati S, and Baumgart-Vogt E. Peroxisomes in airway epithelia and future prospects of these organelles for pulmonary cell biology. *Histochem Cell Biol.* 2009; 131(4): 447-454. DOI: [10.1007/s00418-009-0566-4](https://doi.org/10.1007/s00418-009-0566-4)
 92. Estévez M, and Xiong Y. Intake of oxidized proteins and amino acids and causative oxidative stress and disease: Recent scientific evidences

- and hypotheses. J Food Sci. 2019; 84(3): 387-396. DOI: [10.1111/1750-3841.14460](https://doi.org/10.1111/1750-3841.14460)
93. Cheah LT, Hindle MS, Khalil JS, Duval C, Unsworth AJ, and Naseem KM. Cells. 2025; 14(7): 500. DOI: [10.3390/cells14070500](https://doi.org/10.3390/cells14070500)
94. ang SY, and Choi KM. Impact of adipose tissue and lipids on skeletal muscle in sarcopenia. J Cachexia Sarcopenia Muscle. 2025; 16(4): e70000. DOI: [10.1002/jcsm.70000](https://doi.org/10.1002/jcsm.70000)
95. Min L, Cheng J, Zhao S, Tian H, Zhang Y, Li S, et al. Plasma-based proteomics reveals immune response, complement and coagulation cascades pathway shifts in heat-stressed lactating dairy cows. J Proteomics. 2016; 146: 99-108. DOI: [10.1016/j.jprot.2016.06.008](https://doi.org/10.1016/j.jprot.2016.06.008)
96. Pevzner IB, Andrianova NV, Lomakina AK, Cherksova KS, Semenchenko ED, and Plotnikov EY. Organ-specific extracellular vesicles in the treatment of ischemic acute organ injury: Mechanisms, successes, and prospects. Int J Mol Sci. 2025; 26(19): 9709. DOI: [10.3390/ijms26199709](https://doi.org/10.3390/ijms26199709)
97. Akash MSH, Yaqoob A, Rehman K, Imran M, Assiri MA, Al-Rashed F, et al. Metabolomics: A promising tool for deciphering metabolic impairment in heavy metal toxicities. Front Mol Biosci. 2023; 10: 1218497. DOI: [10.3389/fmolb.2023.1218497](https://doi.org/10.3389/fmolb.2023.1218497)
98. Ducret V, Richards AJ, Videlier M, Scalvenzi T, Moore KA, Paszkiewicz K, et al. Transcriptomic analysis of the trade-off between endurance and burst-performance in the frog *Xenopus allofraseri*. BMC Genomics. 2021; 22(1): 204. DOI: [10.1186/s12864-021-07517-1](https://doi.org/10.1186/s12864-021-07517-1)
99. Loor JJ. Genomics of metabolic adaptations in the peripartal cow. Animal. 2010; 4(7): 1110-1139. DOI: [10.1017/S1751731110000960](https://doi.org/10.1017/S1751731110000960)